

## THE DEVELOPMENT OF THE PCR TEST SYSTEM FOR IDENTIFICATION OF RNA-2 OF BNYVV FOR ENVIRONMENTAL ASSESSMENT OF PHYTOSANITARY STATE\*

K. Hrynychuk

*Національний університет біоресурсів і природокористування України*

*Проведено біоінформативний аналіз нуклеотидних послідовностей РНК-2 ізолятів вірусу некротичного пожовтіння жилок буряку (ВНПЖБ). Встановлено консервативні послідовності гена, що кодує протеїн Р75 ВНПЖБ, та розроблено дизайн праймерів, специфічних до нуклеотидних послідовностей геному для ідентифікації протеїну Р75 РНК-2 ВНПЖБ. Синтезовано специфічні олігонуклеотидні праймери Forward 5'-CT-TTGGCAGGATTAGGCTCG -3', Reverse 5'-САСТСГГГАСТАТСАССАГГ -3'. Розроблену систему було випробувано та підтверджено ефективність її використання на моделі кДНК українського ізоляту ВНПЖБ. Оптимізовано систему ідентифікації РНК-2 за температурними показниками відпалу праймерів і встановлено, що оптимальна температура варіює в межах 50–62°C. Отримані результати дають змогу ідентифікувати РНК-2 ВНПЖБ у зразках цукрових буряків та створюють передумови для розробки кількісних методів діагностики та ідентифікації ВНПЖБ.*

**Ключові слова:** вірус некротичного пожовтіння жилок буряку (ВНПЖБ), РНК-2, ген, ПЛР, дизайн, праймер, ідентифікація.

Beet necrotic yellow vein virus (BNYVV) is the type member of the Benyvirus genus, the causal agent of sugarbeet rhizomania. Rhizomania disease of sugar beet was first reported in Italy in 1952 by Canova. BNYVV has been reported in Europe (Austria, Belgium, Bulgaria, France, Germany, Iran, Hungary,

Greece, Czech Republic, Kazakhstan, Kyrgyzstan, Netherlands, Poland, Romania, Russia, Slovakia, Spain, Sweden, Switzerland, Yugoslavia, Great Britain, Asia (China, Japan, Syria) and North America [1–4]. In Ukraine BNYVV is included in the register of quarantine viruses.

Genome of BNYVV consists of 4–5 particles of (+) RNA. The RNA-2, 4612 nucleotide in length [excl. the poly (A)-tail], has 6 open reading frames (ORF) which code genes of encapsidation, cell-to-cell movement and suppression of posttranscriptional gene silencing (PTGS). RNA-2 is necessary for

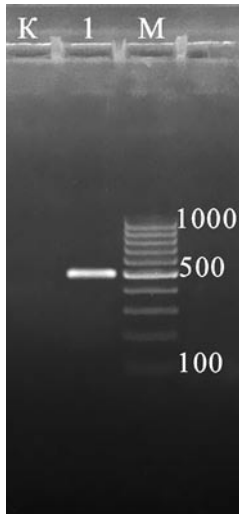
replication. The first 5' ORF codes for a 21 kDa CP with rather weak UAG termination codon [5]. RNA-2 codes triple gene block (TGB)-encoded proteins (TGBp1, TGBp2, TGBp3) which is necessary for cell-to-cell movement. Cysteine-rich protein P14 is expressed by subgenomic RNA [6]. Protein P75 is involved in vector transmission by *Polytuha betae* [7].

Since the infection of sugar beet with BNYVV decreases the yield, it is important to prevent the spread of the virus, so it is necessary to carry out timely virus identification. The aim of our research was to develop diagnostic system of BNYVV based on polymerase chain reaction for the identification protein P75 of RNA-2.

### MATERIALS AND METHODS

For searching nucleotide sequences of RNA-2 and for bioinformative analysis a database of NCBI (National Center for Biotechnological Information) was used [8]. The bioinformatics genome analysis was performed using the software «MultAlin» (Multiple se-





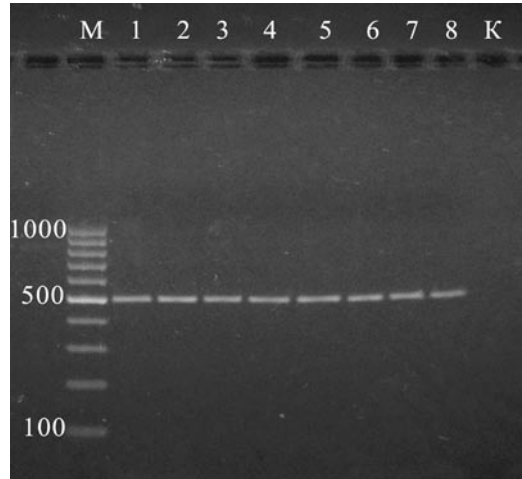
**Fig. 2.** The electrophoregram of PCR product analysis for the identification of RNA-2 of BNYVV: K – negative control of PCR analysis, 1 – amplification product of P75 gene, M (GeneRuler 10 bp DNA Lader 0241) – marker of fragment lengths (base pairs)

The PCR was performed using cDNA of Ukrainian isolate of BNYVV under the following conditions: initial denaturation 5 min – 94°C; 30 cycles: denaturation 30 sec – 94°C, annealing of primers 30 sec – 60°C, elongation 30 sec – 72°C, 72°C final synthesis – 7 min.

As a result of the visualization of PCR amplification product fragments of size – 490 nucleotides pairs were revealed. An availability on the electrophoregram expected reaction product indicates that the developed system for the identification of RNA-2 of BNYVV is effective (Fig. 2).

On the next stage of the study we had to pick up the operating temperature annealing of primers. We made a series of reactions with temperature annealing of primers from 50°C to 64°C. PCR was performed under the following conditions: initial denaturation of 5 min – 94°C; 20 cycles: denaturation 30 sec – 94°C annealing of primers with 30 sec – 50–64°C, 30 sec elongation – 72°C, 72°C final synthesis – 7 min.

Although primers annealing temperature conditions did not significantly affect on the efficiency of amplification reactions, we have found that more optimal temperature is in the range of 50–62°C. Under these conditions nonspecific amplification products were not observed and the amount of amplicons was sufficient for clear visualization in agarose gel (Fig. 3).



**Fig. 3.** The electrophoregram of the temperature optimization of primers annealing for the identification of gene P75: M (GeneRuler 10 bp DNA Lader 0241), 1 – 50°C, 2 – 52°C, 3 – 54°C, 4 – 56°C, 5 – 58°C, 6 – 60°C, 7 – 62°C, 8 – 64°C, K – negative control of PCR analysis

## CONCLUSIONS

a) A bioinformative analysis of nucleotide sequences of RNA-2 of world isolates of BNYVV was carried out. Conserved sequences of the gene encoding protein P75 of BNYVV were shown and implemented design primers for identification of RNA-2 of BNYVV was made. The primers with optimal characteristics such as: Forward 5'-CTTTGGCAGGATTAGGCTCG-3', Reverse 5'-CACTCGGGACTATCACCAGG-3' were synthesized.

b) The developed system was tested and confirmed the effectiveness of its use on Ukrainian model cDNA isolate of BNYVV.

c) The annealing temperature of primers was optimized. The optimal temperature for identification of BNYVV was in the range of 50–62°C.

In our further studies we are planning to determine the nucleotide sequences of RNA-2 of P75 protein and establish phylogenetic relationship with world isolates.

## REFERENCES

1. Nucleotide Sequence of Beet Necrotic Yellow Vein Virus RNA-2 / [S. Bouzoubaa, V. Ziegler, D. Beck et al.]. // *J. gen. Virol.* – 1986. – No. 67. – P. 1689–1700.
2. Pavli O.I. Molecular characterization of beef necrotic yellow vein virus in Greece and transgenic approaches towards enhancing rhizomania disease resistance: dissert. of doctor of agricultural sciences / O.I. Pavli. – Wageningen, 2010. – 166 p.
3. *Pferdmenges F.* Occurrence, spread and pathogenicity of different Beet necrotic yellow vein virus (BNYVV) isolates: dissertation zur Erlangung des Doktorgrades der Fakultät für Agrarwissenschaften / Pferdmenges Friederike. – Göttingen, 2007. – 111 s.
4. Canova A. Appunti di patologia della barbabietola / A. Canova // *Inf. Fitopatol.* – 1959. – No. 20 – P. 390–396.
5. Virus Taxonomy: The Classification and Nomenclature of Viruses. Ninth Report of the International Committee on Taxonomy of Viruses / [Andrew M.Q. King, Michael J. Adams, Eric B. Carstens et al.]. – San Diego, CA. USA: Elsevier Academic Press, 2011. – 1327 p.
6. Identification, subcellular localization and some properties of a cysteine-rich suppressor of gene silencing encoded by peanut clump virus / [P. Dunoyer, S. Pfeffer, C. Fritsch et al.]. // *Plant Journal.* – 2002. – No. 29. – P. 555–567.
7. Tamada T. Evidence that the 75K readthrough protein of beet necrotic yellow vein virus RNA-2 is essential for transmission by the fungus *Polymyxa betae* / T. Tamada, T. Kusume // *Journal of General Virology.* – 1991. – No. 72. – P. 1497–1504.
8. National Center for Biotechnology Information [Электронный ресурс]. – Режим доступа до ресурсу: <http://www.ncbi.nlm.nih.gov/pubmed>
9. Multiple sequence alignment by Florence Corpet [Электронный ресурс]. – Режим доступа до ресурсу: <http://multalin.toulouse.inra.fr/multalin>
10. Primer3web version 4.0.0 – Pick primers from a DNA sequence [Электронный ресурс]. – Режим доступа: <http://primer3.ut.ee>

## REFERENCES

1. Bouzoubaa S., Ziegler V., Beck D. et al. (1986). Nucleotide Sequence of Beet Necrotic Yellow Vein Virus RNA-2, *J. gen. Virol.* no. 67, pp. 1689–1700 (*in English*).
2. Pavli O.I. (2010). Molecular characterization of beef necrotic yellow vein virus in Greece and transgenic approaches towards enhancing rhizomania disease resistance: dissert. of doctor of agricultural sciences, Wageningen, 166 p. (*in English*).
3. *Pferdmenges F.* (2007). Occurrence, spread and pathogenicity of different Beet necrotic yellow vein virus (BNYVV) isolates: dissertation zur Erlangung des Doktorgrades der Fakultät für Agrarwissenschaften Göttingen, 111 p. (*in English*).
4. Canova A. (1959). Appunti di patologia della barbabietola. *Inf. Fitopatol.* no. 20, pp. 390–396 (*in English*).
5. Andrew M.Q. King, Michael J. Adams, Eric B. Carstens (2011). Virus Taxonomy: The Classification and Nomenclature of Viruses. Ninth Report of the International Committee on Taxonomy of Viruses. San Diego, CA. USA: Elsevier Academic Press, 1327 p. (*in English*).
6. Dunoyer P., Pfeffer S., Fritsch C. (2002). Identification, subcellular localization and some properties of a cysteine-rich suppressor of gene silencing encoded by peanut clump virus. *Plant Journal*, no. 29, pp. 555–567 (*in English*).
7. Tamada T., Kusume T. (1991). Evidence that the 75K readthrough protein of beet necrotic yellow vein virus RNA-2 is essential for transmission by the fungus *Polymyxa betae*. *Journal of General Virology*, no. 72, pp. 1497–1504. (*in English*).
8. National Center for Biotechnology Information. Available at: <http://www.ncbi.nlm.nih.gov/pubmed> (*in English*).
9. Multiple sequence alignment by Florence Corpet. Available at: <http://multalin.toulouse.inra.fr/multalin> (*in English*).
10. Primer3web version 4.0.0 – Pick primers from a DNA sequence. Available at: <http://primer3.ut.ee> (*in English*).